An MRI study of age-related white and gray matter volume changes in the rhesus monkey

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Abstract

We applied the automated MRI segmentation technique Template Driven Segmentation (TDS) to dual-echo spin echo (DE SE) images of eight young (5–12 years), six middle-aged (16–19 years) and eight old (24–30 years) rhesus monkeys. We analyzed standardized mean volumes for 18 anatomically defined regions of interest (ROI’s) and found an overall decrease from young to old age in the total forebrain (5.01%), forebrain parenchyma (5.24%), forebrain white matter (11.53%), forebrain gray matter (2.08%), caudate nucleus (11.79%) and globus pallidus (18.26%). Corresponding behavioral data for five of the young, five of the middle-aged and seven of the old subjects on the Delayed Non-matching to Sample (DNMS) task, the Delayed-recognition Span Task (DRST) and the Cognitive Impairment Index (CII) were also analyzed. We found that none of the cognitive measures were related to ROI volume changes in our sample size of monkeys.

Keywords: Aging; Rhesus monkey; MRI; White matter; Gray matter; Cerebral cortex; Template Driven Segmentation (TDS)

1. Introduction

In selecting an appropriate animal model in which to study the effects of normal aging, the most important criteria that must be met is how closely the animal model reflects human neurobiology. The rhesus monkey (Macaca mulatta) qualifies as an excellent non-human primate model for the study of normal aging for several reasons. First, the anatomy of the brain more closely resembles that of the human brain than other available laboratory animals. Second, unlike the human brain, the rhesus monkey brain does not exhibit the characteristic neuropathological loss of neurons that characterizes Alzheimer’s disease (Peters et al., 1998) and hence normal aging can be studied without risk of intrusion of AD cases into the sample. Third, the life span of the human compared to the rhesus monkey approximately follows a ratio of 3:1 (Bowden and Williams, 1984; Tigges et al., 1988); Based on this, monkeys age 5–12 years are considered “young adults”, those between 13 and 19 are considered middle aged and those beyond 20 years are considered “aged adults” (Peters et al., 1996). Fourth, monkeys can be evaluated on batteries of behavioral tasks that tap cognitive domains that clearly correspond to human cognitive functions (Herndon et al., 1997). Fifth, monkeys and humans show similar patterns of age-related cognitive changes (Moss and Albert, 1988).
Studies of human aging post-mortem have been faced with the difficult problem of obtaining optimally prepared brain tissue from behaviorally well-characterized subjects. Recent advances in MRI technology, however, have allowed in vivo anatomical assessments to be conducted with closely monitored human subjects. These studies of the human brain have reported a variety of age-related changes in the volumes of different tissue components and have reported loss of gray matter, loss of white matter and loss of both (Bartzokis et al., 2001; Condon et al., 1986; Ge et al., 2002; Gunning-Dixon et al., 1998; Guttmann et al., 1998; Jernigan et al., 1991; Kettenen, 1998; Krishnan et al., 1990; McDonald et al., 1991; Meier-Rüge et al., 1992; Raz et al., 1995, 1997, 2003, 2004; Resnick et al., 2003; Salat et al., 1999). For example, using manual tracing methods, Raz et al. (1997, 2004) reported gray matter and white matter volume decrease in the prefrontal cortex and medial temporal lobe from high resolution spoiled gradient recalled (SPGR) images and Bartzokis et al. (2001) reported gray matter volume loss in the frontal and temporal lobes accompanied by a quadratic (increase then decrease) change of white matter volume with increasing age in the same lobes from dual-echo spin echo images. However, using automated segmentation methods, Guttmann et al. (1998) reported white matter volume decrease, with a minor change in gray matter volume on dual-echo spin echo images, and Ge et al. (2002) reported total gray and white matter volume loss with age in both men and women on dual-echo fast spin echo images.

Studies involving the rhesus monkey as an animal model for normal aging have focused on behavioral (Bachevalier and Mishkin, 1989; Bachevalier et al., 1991; Herndon et al., 1997; Lai et al., 1995; Mahut et al., 1982; Mishkin, 1978; Moore et al., 2003, 2005; Moss et al., 1988, 1997; Peters et al., 1996, 1998; Peters, 1999; Presty et al., 1987; Rapp and Amaral, 1989, 1991) and anatomical (Andersen et al., 1999; Matochik et al., 2000) endpoints, but few have examined both (Peters et al., 1996, 1998; Peters, 1999). The rhesus monkey model provides the opportunity to study age-related brain structural changes using MRI with the assurance that Alzheimer’s disease pathology will not confound the interpretation of results. Unfortunately unlike human aging studies, only a handful of rhesus monkey studies have utilized structural MRI analysis for the study of aging.

For example, Andersen et al. (1999) used a signal intensity classification algorithm to segment female monkey brains into gray matter, white matter and cerebrospinal fluid (CSF) classes. They found a decrease in parenchyma volume (normalized to the intracranial cavity volume). Andersen and colleagues attributed the decline in parenchyma volume to a decrease in gray matter volume and a compensatory increase in CSF volume, with some white matter volume loss up to 15 years of age. However, for later years of the monkeys’ lives, they attributed parenchyma volume loss mostly to white matter volume loss.

In another example, Matochik et al. (2000) used a manual tracing segmentation method to measure the volume of the rhesus monkey striatum. In 19 male monkeys between the ages of 3 and 30 years, they found age-related declines in normalized caudate nucleus and putamen volumes when comparing young, middle-aged and aged groups. Matochik and colleagues postulated that the volume loss could be due to both neuronal- and non-neuronal-related atrophy (Matochik et al., 2000; Morris et al., 1999).

The rhesus monkey model of normal aging also provides the opportunity to study cognitive decline (Bachevalier and Mishkin, 1989; Bachevalier et al., 1991; Herndon et al., 1997; Lai et al., 1995; Mahut et al., 1982; Mishkin, 1978; Moore et al., 2003, 2005; Moss et al., 1988, 1997; Peters et al., 1996, 1998; Peters, 1999; Presty et al., 1987; Rapp and Amaral, 1989, 1991) using behavioral tasks that are similar to clinical tests (Barbeau et al., 2005; Dickerson et al., 2004; Li et al., 2004; Pfefferbaum et al., 2001). Similar to anatomical studies, the advantage of using the rhesus monkey as an animal model for cognitive decline in normal aging is the absence of Alzheimer’s disease-associated cognitive impairment.

The goal of this investigation was to utilize MRI methods to determine if there are age-related changes in the major components of the forebrain in behaviorally characterized monkeys participating in ongoing investigations of normal aging (Herndon et al., 1997; Lai et al., 1995; Moore et al., 2003, 2005; Moss et al., 1988, 1997; Peters et al., 1996, 1998; Peters, 1999). To accomplish this efficiently we applied the automated MRI segmentation technique Template Driven Segmentation (TDS) to legacy rhesus monkey MRI data. The technique has been validated in human studies of normal aging (Guttmann et al., 1999; Iosifescu et al., 1997; Warfield et al., 1995; Wei et al., 2002). We created a rhesus monkey anatomical atlas template for the segmentation pipeline to ensure that all voxels were assigned to anatomically validated components. Age-related volume changes calculated by TDS were then compared to cognitive performance as measured by the Delayed Non-matching to Sample (DNMS), the Delayed Recognition Span Task (DRST) and the Cognitive Impairment Index (CII), which is a composite score of DNMS acquisition, DNMS 2-min delay and DRST spatial condition tasks. The underlying hypothesis for this investigation was that age-related brain structure volume decreases would be associated with impaired cognitive performance.

2. Materials and methods

2.1. Selection of animal subjects and housing accommodations

A total of 22 male rhesus monkeys were selected for this study, 8 of whom were young (5–12 years), 6 of whom were middle-aged (16–19 years) and 8 of whom were old (24–30 years) at the time of MRI scan acquisition. All subjects were selected from the rhesus monkey population at the Yerkes...
National Primate Research Center (YNPRC) according to explicit criteria which excluded subjects with histories that included any of the following: splenectomy or thymectomy, exposure to radiation, organ transplantation, malnutrition, cancer, chronic illness, chronic drug administration or any neurological disease. Prior to entry into the study, all animals received a medical examination that included serum chemistry, hematology, urine analysis and fecal analysis as well as assessment of visual function to ensure that behavioral testing would not be impeded. Once accepted, quarterly physicals were conducted to ensure continued health.

While participating in the study, all monkeys were housed first at YNPRC and subsequently transferred to the Laboratory Animal Science Center at Boston University Medical Center (BUMC) where they were individually housed within auditory and visual range of other monkeys. They were all maintained on a normal diet consisting of Standard Purina Monkey Chow and under a 12-h light/dark cycle consisting of a gradual transition of 1 h. Feeding occurred once a day following behavioral testing, and water was available continually. The monkeys were also inspected visually on a daily basis by both animal care personnel and research technicians. All procedures were approved by the Institutional Animal Care and Use Committees (IACUC) of both YNPRC and BUMC. In addition both YNPRC and BUMC are fully accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care (AAALAC) for distinguishing low contrast brain structures on the DE SE images to improve signal-to-noise ratio while preserving structure edges (Gerig et al., 1992).

2.3. Creation of a reference atlas

A reference label atlas of the monkey brain was constructed by manual outlining of DE SE images of AM093 using 3D Slicer (http://www.slicer.org) after anisotropic diffusion filtering. The resulting atlas consisted of 14 manually segmented ROI’s: forebrain white matter, cerebral cortex, caudate nucleus, putamen, globus pallidus, claustrum, thalamus, hypothalamus, brainstem, cerebellum, lateral ventricles, third ventricle, cerebral aqueduct and fourth ventricle. Operational definitions for the segmented ROI’s can be found in Appendix A.

Although the pdw images provided good contrast between gray matter and white matter regions, and the T2w images provided good contrast between brain and CSF, Nissl stained histological sections from AM093, digitized using the Inquiry digitizing system (Loats Associates, Inc.), and a published histological atlas (Paxinos et al., 1999), served as anatomical references for correctly labeling each ROI and for distinguishing low contrast brain structures on the DE SE images. The atlas was validated by four additional independent raters who identified the same 14 ROI’s on the DE SE images of the atlas subject (P = 0.4664, repeated measures ANOVA).

An intracranial cavity (ICC) ROI (Guttmann et al., 1998; Kikinis et al., 1992; Warfield et al., 1995; Wei et al., 2002) was created using the label threshold function in 3D Slicer and included the entire cerebrum, cerebellum and brainstem (up to the most inferior margin of the cerebellum). ICC volumes were not significantly different between age groups [F(2,19) = 0.79, P = 0.4699]. The ICC ROI volume was used to standardize all other ROI volumes. The ICC ROI was also used as the reference matrix for the first of the two registration steps in the non-linear registration algorithm described subsequently.

2.4. Segmentation procedure for subjects

DE SE brain images of all 22 monkeys were segmented using an automated pipeline with minimal rater intervention (Fig. 1). First, anisotropic diffusion filtering smoothed the images. Second, an intracranial cavity (ICC) ROI (Guttmann et al., 1998; Kikinis et al., 1992; Warfield et al., 1995; Wei et al., 2002) was created. Third, a data set statistically segmented for gray matter, white matter and cerebrospinal fluid (CSF) was generated using the 3D Expectation-Maximization (3D EM) algorithm, which has been described previously (Wells et al., 1994, 1996). Fourth, Template Driven Segmentation (TDS) (Guttmann et al., 1999; Iosifescu et al., 1997; Warfield et al., 1995; Wei et
Fig. 1. Automated segmentation procedure. Interleaved proton density weighted (pdw) and T2-weighted images (T2w) served as the input for the segmentation pipeline. An intracranial cavity mask (ICC) was created, separating “brain” from “non-brain voxels.” Brain voxels were segmented into gray matter, white matter and CSF classes using the 3D Expectation-Maximization (3D EM) algorithm. An <i>a priori</i> atlas (Atlas) containing 14 regions of interest (ROI’s) segmented from a dual-echo spin-echo (DE SE) data set of a young (7 years old), behaviorally normal monkey (AM093) was created and then elastically matched to the 3D EM results to produce an anatomically based segmentation (TDS) of the gray matter, white matter and CSF voxels. The ROI’s included forebrain white matter, cerebral cortex, caudate nucleus, putamen, globus pallidus, claustrum, thalamus, hypothalamus, brainstem, cerebellum, lateral ventricles, third ventricle, cerebral aqueduct and fourth ventricle, whose operational definitions are made explicit in Appendix A. Colors for “3D EM”: gray = gray matter; white = white matter; blue = CSF. Colors for “Atlas” and “TDS”: gray = cortical gray matter; white = forebrain white matter; blue = ventricles; green = basal ganglia structures (caudate nucleus, putamen, globus pallidus, claustrum, each analyzed separately); orange = thalamus; purple = brainstem. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of the article.)

al., 2002) parcellated the gray matter and CSF further into anatomically defined regions based on a reference label atlas created specifically for this data set.

TDS accomplished the anatomical segmentation by first performing a linear transformation of the reference ICC to the subject ICC (for this step of the segmentation pipeline, the reference data set was the input, and the subject data sets were the target). The resulting transformation matrix was then applied to the reference atlas so that it was now also linearly registered with the subject ICC and 3D EM segmented data set. Finally, TDS performed a non-linear, elastic matching registration of the reference atlas to the 3D EM segmented data set to anatomically classify each voxel. This pipeline was repeated for each subject.

The results provided further segmentation of the gray matter, white matter and CSF regions into more specific regions of interest (ROI) contained in the template atlas. ROI’s of total forebrain (sum of white matter, gray matter and ventricles, but not subarachnoid CSF around the brain or in sulci, fissures and cisterns), forebrain parenchyma (sum of white matter and gray matter), forebrain gray matter (sum of cerebral cortex, caudate nucleus, putamen, globus pallidus, claustrum, thalamus, hypothalamus), and ventricular cerebrospinal fluid (sum of lateral ventricles, third ventricle, cerebral aqueduct and fourth ventricle) were derived.

2.5. Behavioral testing

Five of the young, five of the middle-aged and seven of the old subjects whose brains were imaged also had behavioral testing data available from a battery of cognitive tasks assessing learning and memory as part of ongoing studies. We matched the behavioral data as close to the date of imaging as possible, resulting in a mean date difference of 1 year for young and middle-aged monkeys and 3 years for old monkeys. For all three age groups, the date difference was within the group age range (i.e. behavioral data for old monkeys was not acquired while they were middle-aged). A summary of behavioral tasks is shown in Table 1.

The tasks included the Delayed Non-matching to Sample (DNMS) task (acquisition, 2-min delay and 10-min delay conditions) and the Delayed Recognition Span Task (DRST) (spatial and object conditions). The details of these tasks and the changes associated with normal aging in our monkeys
Table 1
Animal subjects age, age at scan and behavioral scores

<table>
<thead>
<tr>
<th>Age category</th>
<th>Age at scan</th>
<th>Age at testing criterion</th>
<th>Behavioral measures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>DNMS acquisition</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>DNMS 2-min (% correct)</td>
</tr>
<tr>
<td>Young</td>
<td>5</td>
<td>7</td>
<td>45</td>
</tr>
<tr>
<td>Young</td>
<td>6</td>
<td>7</td>
<td>62</td>
</tr>
<tr>
<td>Young</td>
<td>7</td>
<td>7</td>
<td>71</td>
</tr>
<tr>
<td>Young</td>
<td>10</td>
<td>8</td>
<td>52</td>
</tr>
<tr>
<td>Young</td>
<td>10</td>
<td>10</td>
<td>60</td>
</tr>
<tr>
<td>Mean (S.E.)</td>
<td>8.38 (0.86)</td>
<td>8.00 (0.55)</td>
<td>58.00 (4.44)</td>
</tr>
<tr>
<td>Middle-aged</td>
<td>16</td>
<td>16</td>
<td>6</td>
</tr>
<tr>
<td>Middle-aged</td>
<td>16</td>
<td>16</td>
<td>238</td>
</tr>
<tr>
<td>Middle-aged</td>
<td>17</td>
<td>17</td>
<td>114</td>
</tr>
<tr>
<td>Middle-aged</td>
<td>17</td>
<td>17</td>
<td>16</td>
</tr>
<tr>
<td>Middle-aged</td>
<td>19</td>
<td>19</td>
<td>97</td>
</tr>
<tr>
<td>Mean (S.E.)</td>
<td>16.83 (0.48)</td>
<td>17.00 (0.55)</td>
<td>94.20 (41.82)</td>
</tr>
<tr>
<td>Old</td>
<td>24</td>
<td>22</td>
<td>151</td>
</tr>
<tr>
<td>Old</td>
<td>24</td>
<td>21</td>
<td>77</td>
</tr>
<tr>
<td>Old</td>
<td>25</td>
<td>21</td>
<td>60</td>
</tr>
<tr>
<td>Old</td>
<td>27</td>
<td>24</td>
<td>353</td>
</tr>
<tr>
<td>Old</td>
<td>30</td>
<td>29</td>
<td>70</td>
</tr>
<tr>
<td>Old</td>
<td>30</td>
<td>30</td>
<td>29</td>
</tr>
<tr>
<td>Old</td>
<td>30</td>
<td>26</td>
<td>98</td>
</tr>
<tr>
<td>Mean (S.E.)</td>
<td>27.50 (1.00)</td>
<td>24.71 (1.41)</td>
<td>119.71 (41.39)</td>
</tr>
</tbody>
</table>

Mean (S.E.) are summarized at the bottom of each age group. Abbreviations: Delayed Non-matching to Sample (DNMS), Delayed Recognition Span Task (DRST), Cognitive Impairment Index (CII).
have been presented in detail in Herndon et al. (1997) and will only be summarized briefly here.

The tasks were given to all subjects in a fixed order beginning with the acquisition condition of the DNMS task. In the acquisition phase there was a 10 s delay between the presentation of a trial unique sample stimulus and a choice trial where the sample object was paired with a novel object. This assessed the monkey’s ability to learn the non-match rule in which the novel object must be identified and chosen to receive a reward. After learning was achieved by performing at a criterion of 90% correct over 100 trials, the DNMS delay phase was given to assess recognition memory over time. In this phase animals were tested for 100 trials with a delay between sample object presentation and choice trial of 120 s (2 min) and this was followed by another 100 trials with a delay of 600 s (10 min).

After completion of DNMS training and testing, animals were tested on the Delayed Recognition Span Task (DRST), which assessed the animals’ working memory capacity in both the spatial and object modalities. For the spatial DRST condition, identical black plaques were used as stimuli on an 18 well board. On the first trial one plaque was presented in

Fig. 2. Standardized ROI volume for total forebrain (a), forebrain parenchyma (b), forebrain white matter (c), forebrain gray matter (d), caudate nucleus (e) and globus pallidus (f) as a function of age category. Bars and $P$-values indicate significant changes between cohorts.
1 of the 18 locations and covered the reward. On the second trial, with the location of the first plaque unchanged, a new plaque was added over a new baited location and the monkey must use the non-match rule to pick the new plaque to obtain a reward. On subsequent trials, new plaques were added over a new location until an error was made. In the object condition of the task, the same novelty rule applied as new objects were sequentially added, but the spatial position of all objects on the board varied from trial to trial so that object features are the only cue. The number of objects was increased trial to trial until an error was made.

We also calculated the Cognitive Impairment Index (CII), which was a composite score of DNMS acquisition, DNMS 2-min delay and DRST spatial condition tasks. Higher CII scores indicated greater cognitive impairment.

2.6. Data analysis

For each subject, ROI volumes were normalized by dividing the ROI volume by the ICC volume. These normalized volumes were used for all statistical analyses such that “ROI volume” always refers to a standardized measure.

We analyzed the relationship of each ROI volume (total forebrain, forebrain parenchyma, forebrain white matter, forebrain gray matter, cerebral cortex, caudate nucleus, putamen, globus pallidus, claustrum, thalamus, hypothalamus, brainstem, cerebellum, ventricular cerebrospinal fluid, lateral ventricles, third ventricle, cerebral aqueduct and fourth ventricle) with age as a between groups factor for one-way ANOVA, then followed up with Bonferroni a posteriori comparisons.

We also analyzed the relationship of those ROI volumes shown to be significantly associated with age group with individual performance on the DNMS task, DRST and the CII using multiple regression. For these analyses, performance on each behavioral task was the outcome variable and ROI volume and age (linear and non-linear) were predictor variables. All calculations were performed by the statistical software package, Stata 8.2 (Stata Corp, College Station, TX).

3. Results

3.1. ROI volume as a function of age group

One-way ANOVA analysis revealed a significant overall main effect of age group for the following standardized ROI volumes (Fig. 2): total forebrain \( F(2,19) = 20.76, P < 0.0001 \), forebrain parenchyma \( F(2,19) = 20.40, P < 0.0001 \), forebrain white matter \( F(2,19) = 12.89, P = 0.0003 \), forebrain gray matter \( F(2,19) = 3.96, P = 0.0365 \), caudate nucleus \( F(2,19) = 11.48, P = 0.0005 \), globus pallidus \( F(2,19) = 5.39, P = 0.0140 \) and third ventricle \( F(2,19) = 4.53, P = 0.0247 \), data not shown. Cerebral cortex \( F(2,19) = 0.76, P = 0.4800 \) and putamen \( F(2,19) = 1.40, P = 0.2700 \) volumes were not found to be significantly different between age groups.

### Table 2
Mean (S.E.) of standardized ROI volumes according to age group

<table>
<thead>
<tr>
<th>Age category</th>
<th>Regions of interest (ROI)</th>
<th>Total forebrain*#</th>
<th>Forebrain parenchyma*#</th>
<th>Forebrain white matter*#</th>
<th>Forebrain gray matter#</th>
<th>Cerebral cortex</th>
</tr>
</thead>
<tbody>
<tr>
<td>Young</td>
<td></td>
<td>80.61 (0.64)</td>
<td>78.54 (0.68)</td>
<td>26.25 (0.67)</td>
<td>52.28 (0.53)</td>
<td>41.66 (0.61)</td>
</tr>
<tr>
<td>Middle-Aged</td>
<td></td>
<td>83.06 (0.57)</td>
<td>80.96 (0.59)</td>
<td>27.98 (0.56)</td>
<td>52.98 (0.56)</td>
<td>42.36 (0.44)</td>
</tr>
<tr>
<td>Old</td>
<td></td>
<td>76.57 (0.81)</td>
<td>74.42 (0.80)</td>
<td>23.23 (0.68)</td>
<td>51.19 (0.21)</td>
<td>41.58 (0.25)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Age category</th>
<th>Regions of interest (ROI)</th>
<th>Caudate nucleus+#</th>
<th>Putamen</th>
<th>Globus pallidus#</th>
<th>Claustrum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Young</td>
<td></td>
<td>2.29 (0.13)</td>
<td>2.65 (0.14)</td>
<td>1.15 (0.09)</td>
<td>0.54 (0.01)</td>
</tr>
<tr>
<td>Middle-aged</td>
<td></td>
<td>2.81 (0.08)</td>
<td>2.41 (0.16)</td>
<td>1.29 (0.04)</td>
<td>0.50 (0.03)</td>
</tr>
<tr>
<td>Old</td>
<td></td>
<td>2.02 (0.11)</td>
<td>2.34 (0.12)</td>
<td>0.94 (0.07)</td>
<td>0.57 (0.02)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Age category</th>
<th>Regions of interest (ROI)</th>
<th>Thalamus</th>
<th>Hypothalamus</th>
<th>Brainstem</th>
<th>Cerebellum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Young</td>
<td></td>
<td>3.81 (0.29)</td>
<td>0.18 (0.03)</td>
<td>3.40 (0.16)</td>
<td>9.56 (0.41)</td>
</tr>
<tr>
<td>Middle-aged</td>
<td></td>
<td>3.43 (0.21)</td>
<td>0.18 (0.02)</td>
<td>3.26 (0.31)</td>
<td>9.65 (0.55)</td>
</tr>
<tr>
<td>Old</td>
<td></td>
<td>3.52 (0.15)</td>
<td>0.23 (0.02)</td>
<td>3.27 (0.09)</td>
<td>8.74 (0.43)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Age category</th>
<th>Regions of interest (ROI)</th>
<th>Ventricular CSF</th>
<th>Lateral ventricles</th>
<th>Third ventricle</th>
<th>Cerebral aqueduct</th>
<th>Fourth ventricle</th>
</tr>
</thead>
<tbody>
<tr>
<td>Young</td>
<td></td>
<td>2.08 (0.08)</td>
<td>1.73 (0.07)</td>
<td>0.12 (0.02)</td>
<td>0.06 (0.00)</td>
<td>0.16 (0.01)</td>
</tr>
<tr>
<td>Middle-aged</td>
<td></td>
<td>2.10 (0.06)</td>
<td>1.80 (0.06)</td>
<td>0.08 (0.01)</td>
<td>0.06 (0.02)</td>
<td>0.15 (0.02)</td>
</tr>
<tr>
<td>Old</td>
<td></td>
<td>2.15 (0.06)</td>
<td>1.79 (0.07)</td>
<td>0.17 (0.02)</td>
<td>0.06 (0.01)</td>
<td>0.14 (0.01)</td>
</tr>
</tbody>
</table>

Significant volume changes between young and old age (*), between middle age and old age (#) and between young and middle age (+) are indicated with the ROI name.
4.1. Summary of results

We have reported two principal results. First, there was a significant age-related decrease in the ROI volumes of total forebrain, forebrain parenchyma, forebrain white matter, forebrain gray matter, caudate nucleus and globus pallidus. We also found a significant increase in third ventricle volume, but the volume changes were less than 0.2 cm³. Volume changes this small could be accounted for by partial volume effects, and so further analysis was not done for this ROI. Second, none of the ROI volumes significant with age group were associated with cognitive performance on the Delayed Non-matching to Sample (DNMS) task, Delayed Recognition Span Task (DRST) or the Cognitive Impairment Index (CII).

4.2. Volume changes

To our knowledge, the present study uniquely examines ROI volume changes of the entire rhesus monkey forebrain, brainstem and cerebellum as a function of age group. The overall decrease from young to old age in the standardized volumes of total forebrain (5.01%), forebrain parenchyma (5.24%), forebrain white matter (11.53%), forebrain gray matter (2.08%), caudate nucleus (11.79%) and globus pallidus (18.26%) that we found have been similarly reported by many human aging studies (Andersen et al., 1999; Bartokokis et al., 2001; Ge et al., 2002; Gunning-Dixon et al., 1998; Gutmann et al., 1998; Jernigan et al., 1991; Ketonen, 1998; Kohn et al., 1991; Krishnan et al., 1990; Matochik et al., 2000; Meier-Ruge et al., 1992; Murphy et al., 1992; Raz et al., 1995, 2003; Resnick et al., 2003; Salat et al., 1999).

Forebrain white matter and forebrain gray matter volume changes in the monkey closely matches the results of Gutmann et al. (1998) and Ge et al. (2002) in humans. We were surprised, however, to find that ventricular CSF volume did not increase significantly with age since the phenomenon has been established as a marker for normal aging in humans (Pfefferbaum et al., 1994; Tanna et al., 1991). CSF volume increase has also been characterized in the rhesus monkey by Andersen et al. (1999), but their measure included both cortical surface CSF and ventricular CSF. In our sample size, ventricular CSF accounted for only 2.6% of the total brain volume and increased 3% (0.1% of ICC) from young to old age. With a 5% decrease in total brain volume (4% of ICC), only cortical surface CSF increases could balance the loss of brain parenchyma within the ICC. Unfortunately in the present study, we only included ventricular CSF in the template atlas, concerned with the possibility that sulcal CSF would not segment correctly due to partial volume effects. We will need to revisit this issue in order to make a more accurate assessment of total CSF volume changes. Another possibility for the discrepancy between our results and the human data concerns the sample size of eight young, six middle-aged and eight old monkeys. Our small sample size may not have enough statistical power to detect small changes in rhesus monkey ventricular volume.

Although we found a decrease in cerebral cortical volume as a function of age group, that change was not statistically significant ($P = 0.4800$). Cortical volume loss has been reported in human studies, however (Bartokokis et al., 2001; Raz et al., 1997, 2004; Salat et al., 1999), whereas the studies by Bartokokis and colleagues and Raz and colleagues focused on regional areas of cerebral cortex (notably frontal and temporal lobes), our study examined the entire cerebral cortex. It is possible that, when analyzed together, significant decreases in the cortical volume of selective areas are washed out by other areas of the cortex that experience little or no changes in volume. Indeed, Salat et al. (2004) found non-significant differences in global cortical thinning, but follow-up regional analysis of the primary and secondary cortical areas showed significant differences between young and old subjects. Had
we parcellated our template atlas to include regional areas of cerebral cortex, we might have found similar results that have been reported by Bartzokis, Raz and their colleagues.

The overall decrease in caudate nucleus and globus pallidus agrees in part with the results reported in humans (Gunning-Dixon et al., 1998; Murphy et al., 1992; Raz et al., 1995, 2003) and in monkeys (Matochik et al., 2000). The basal ganglia have extensive connections with the prefrontal, temporal and parietal association cortices as well as the thalamus (Graybiel, 2000, 2004; Yeterian and Pandya, 1993, 1998). To determine whether volume changes in forebrain and basal ganglia regions could be age-related, we performed follow-up multiple regression analyses of caudate nucleus and globus pallidus as outcome variables of forebrain white matter, forebrain gray matter (separately) and age (quadratic fit) as predictor variables. Forebrain white matter and forebrain gray matter predicted caudate nucleus volume (but not globus pallidus volume) at trends towards significance of $P = 0.057$, $P = 0.068$, respectively. Again, the age-related relationship between forebrain and basal ganglia ROI volumes may have reached significance had we restricted our analysis to specific lobes. Nevertheless, these results suggest that forebrain and subcortical volumes are age-related. A diffusion tensor imaging study of the subcortical white matter may reveal whether fiber tract integrity also changes with age.

Unlike the study by Matochik et al. (2000), we did not observe a significant decrease in putamen ROI volume even though we were able to corroborate the decrease in caudate nucleus volume. In the present study, we attempted to delineate the claustrum from the lateral edge of the putamen. Partial volume effects at the interface between the two nuclei could easily have affected the calculation of the putamen during the automated segmentation process. Matochik and colleagues do not address whether the claustrum may have influenced the delineation of the putamen. We are not aware of any other rhesus monkey studies of putamen volume that could shed light on the discrepant findings.

We observed an increase in forebrain white matter and forebrain gray matter volume from young to middle age, followed by a decrease from middle age to old age in our sample set of monkeys (Fig. 2), corroborating observations made by Bartzokis et al. (2001) in human subjects. We examined whether this pattern of volume change was true for cerebral cortex, caudate nucleus, putamen, globus pallidus, claustrum, thalamus and hypothalamus ROI’s, the components of the forebrain gray matter ROI. Indeed, the growth pattern was observed for all of the ROI’s except thalamus and hypothalamus, which remained steady in volume through increasing age (data not shown). It is possible that this phenomenon could be caused by a sampling error of larger brains in the middle-aged group, but we verified that the brain weights (measured by water displacement) of the monkeys were only slightly, but not significantly, greater than the monkeys of the other cohorts (Herndon et al., 1997). Another confounding source could be in the segmentation of the tissue classes at the gray/white interface. Since the voxel dimensions in the legacy MRI data were quite large, partial volume effects may have played a role. However, the fact that almost every ROI experienced quadratic volume change leads us to believe that our results may be biological. Bartzokis et al. (2001) attribute the volume changes of white matter relative to gray matter in the frontal and temporal lobes to the timing of myelination and degeneration, respectively, from the second through the fourth decade of life in humans. In post-mortem observations, Kemper (1994) noted an increase in myelination of the association fibers through childhood and adolescence, stabilizing in early middle age, then decreasing in late middle age to old age; Lintl and Braak (1983) reported decreased myelin staining in the visual cortex stripe of Gennari beginning in the third decade of life of normal subjects.

4.3. Cognitive assessment

Changes in total forebrain, forebrain parenchyma, forebrain white matter, forebrain gray matter, caudate nucleus and globus pallidus ROI volumes were not related to cognitive performance on the DNMS task, DRST or the CII. In previous studies, however, we and our colleagues have shown a relationship between cognitive decline on recognition memory tasks (Bachevalier et al., 1991; Herndon et al., 1997; Mahut et al., 1982; Mishkin, 1978; Moss et al., 1988, 1997; Peters et al., 1998; Presty et al., 1987; Rapp and Amaral, 1989, 1991). We believe that the discrepancies between the present and previous results were due to several sources. First, our ROI’s included both hemispheres and spanned the entire forebrain. More specific ROI’s within white matter (and in the gray matter) regions, such as “medial temporal lobe subcortical white matter,” may be related to cognitive performance. However, accurate demarcation of the white matter fibers would require diffusion-weighted data that was not acquired with these monkeys. This endeavor is worth pursuing for future studies. Second, the available behavioral data for the monkeys was incomplete for some tasks, particularly in the middle-aged group. A better-controlled sample size and/or a longitudinal study might be able to provide the necessary statistical power to determine the relationship between cognitive decline on recognition memory tasks and brain volumes.

4.4. Caveats

The demarcation of ROI’s between voxels corresponding to different tissue classes should always be taken with some caution. We readily acknowledge that statistical and anatomical segmentation of brain structures, particularly for the image quality used in our study, will be influenced by partial volume effects.

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Appendix A

The DE SE data were acquired over a 10-year period using a single set of sequence parameters that were optimized at the beginning of the study. Due to the anisotropic voxel dimensions, data were best viewed in the coronal plane. Cerebral cortex, forebrain white matter and lateral ventricles were easily distinguishable in this orientation. However, operationally defined landmarks assisted with the parcellation of the thalamus, hypothalamus, third ventricle, cerebral aqueduct, fourth ventricle, brainstem and cerebellum.

A.1. Operationally defined landmarks

The following landmarks were overlaid on the images of the atlas subject to guide the creation of the ROI’s (Fig. A1).

Fig. A1. Reference landmarks used to operationally define borders of regions of interest (ROI's). The image slices shown here are representative of several slices in which the landmark was used by manual raters. Landmarks: caudate-putamen (a), third ventricle and thalamus–hypothalamus (b), thalamus–brainstem and lateral ventricle (c), brainstem–cerebellum (d) and fourth ventricle (e).
A.1.1. Caudate-putamen landmark
A line parallel to the internal capsule, separating the caudate medially from the putamen laterally where they are connected anteriorly by the nucleus accumbens. The nucleus accumbens is therefore distributed evenly (approximately) between the caudate and putamen.

A.1.2. Third ventricle landmark
An axial line just inferior to the body of the fornix.

A.1.3. Thalamus–hypothalamus landmark
A line that extended from the vertical midpoint of the third ventricle to the most dorsal aspect of the hippocampus. This line was placed on slices where both the thalamus and hypothalamus were adjacent to each other.

A.1.4. Thalamus–brainstem landmark
A line that extended from the most inferior point of the third ventricle to the most dorsal aspect of the hippocampus. This landmark was never present on the same image slice as the thalamus–hypothalamus landmark.

A.1.5. Lateral ventricle landmark
A line tangential to the lateral corner of the lateral geniculate nucleus of the thalamus. This landmark defined the medial border of the inferior horn of the lateral ventricles where the choroidal fissure was difficult to identify.

A.1.6. Brainstem–cerebellum landmark
A parasagittal line along the lateral edge of the brainstem. Bilaterally, these two parallel lines would differentiate the brainstem from the cerebellum on image slices rostral to the appearance of the fourth ventricle such that brainstem is labeled between the lines, and the cerebellum is labeled outside the lines.

A.1.7. Fourth ventricle landmark
A line extended from the lateral corner of the fourth ventricle to the angle between the cerebellum folia and the brainstem. The landmark appeared on the first image slice where the fourth ventricle can be seen between the brainstem and cerebellum.

A.2. Operationally defined structures and regions
The following operational definitions for ROI’s guided the creation of the atlas after operationally defined landmarks were overlaid on the images.

A.2.1. Intracranial cavity (ICC)
Included all pixels of the brain parenchyma and cerebrospinal fluid except the olfactory tract and the optic nerve rostral to the optic chiasm, and the brainstem inferior to the cerebellomedullary cistern.

A.2.2. Total forebrain
Included forebrain white matter, forebrain gray matter and ventricles.

A.2.3. Forebrain parenchyma
Included forebrain white matter and forebrain gray matter.

A.2.4. Forebrain white matter
Included all white matter of the cerebral hemispheres but not white matter contained within the deep gray matter structures of the basal ganglia, brainstem or cerebellum.

A.2.5. Forebrain gray matter
Included the cerebral cortex, caudate nucleus, putamen, globus pallidus, claustrum, thalamus and hypothalamus, but not the brainstem and cerebellum.

A.2.6. Cerebral cortex
Included cerebral cortex, basal forebrain, hippocampus and amygdala, but not nuclei of the deep gray matter.

A.2.7. Caudate nucleus
Striatal gray matter medial to the Caudate-Putamen landmark (including the medial portion of nucleus accumbens) extending throughout the body and tail.

A.2.8. Putamen
Striatal gray matter lateral to the Caudate-Putamen landmark (including the lateral portion of nucleus accumbens).

A.2.9. Globus pallidus
Lenticular nucleus gray matter including internal and external segments, but not including the putamen.

A.2.10. Claustrum
Deep gray matter between putamen and insula surrounding external capsule and extending partially into the temporal stem white matter.

A.2.11. Thalamus
First visible posterior to the optic chiasm, one slice caudal to the first slice on which the hypothalamus first appeared. The thalamus was distinguished from the hypothalamus by the thalamus–hypothalamus landmark. Caudal to the hypothalamus, the thalamus was distinguished from the brainstem by the thalamus–brainstem landmark.

A.2.12. Hypothalamus
First visible on the slice which exhibited the anterior commissure traversing across the midline. On this image the HYP was inferior to the anterior commissure. The thalamus and hypothalamus were distinguished by the thalamus–hypothalamus landmark.
A.2.13. Brainstem

Included all intrinsic white matter and gray matter demarcated by the thalamus–brainstem, brainstem–cerebellum and fourth ventricle landmarks. The brainstem was excluded on images caudal to the section where the cerebellomedullary cistern and fourth ventricle appear continuously.

A.2.14. Cerebellum

Included all intrinsic white matter and gray matter structures demarcated by the brainstem–cerebellum and fourth ventricle landmarks and its interface with subarachnoid CSF and cerebral cortex. The fourth ventricle was not included.

A.2.15. Ventricular CSF

Sum of ventricular volumes.

A.2.16. Lateral ventricles

Included all CSF-associated pixels within the anterior horns, confluence, posterior horns and inferior horns. The lateral ventricle landmark was the medial border for the inferior horns.

A.2.17. Third ventricle

Included all CSF-associated pixels within the diencephalon, extending rostrally between the hemi-structures of the hypothalamus and caudally between the hemispheres of the posterior thalamus up to the transverse fissure. The superior border of the third ventricle was the third ventricle landmark.

A.2.18. Cerebral aqueduct

Included all CSF-associated pixels leading from the third ventricle to the fourth ventricle at the midline of the brainstem.

A.2.19. Fourth ventricle

Included the CSF-associated pixels just caudal to the cerebral aqueduct, between the cerebellum and brainstem as demarcated by the fourth ventricle line, not including the cerebellomedullary cistern.

References


